

I) USING MOLECULAR PHYLOGENETICS TO CHARACTERIZE MICROBIAL BIODIVERSITY.

II) WHY INTERESTED?

III) WHY MICROBES DIFFERENT?

- A) UNCULTURABLE - MANY
- B) PHENOTYPE NOT. U. MEANINGFUL

IV) MOLECULAR PHYLOGENETICS

A) CAN CLONE EVEN IF NOT CULTURE

B) WHAT GENE?

C) WHY rRNA?

- RDP

- 2ARY ALIGN

- PCR PRIMERS

- RIGHT RATE

- SIGNATURE SEQS.

D) HOW

- CLONE - PCR

- SEQUENCE

- ALIGN

- PHYLOGENY = PARSIMONY, DISTANCE, ML

1) Tree of Life

2) 2ary

3) ALIGN w/ PRIMER

4) PCR ✓

5)

V) PARERS

324 3343

328-44

324
8445

PAPERS

I) INTRO ETC.

A) METHODS

① purify ONA

↓
PCR

↓
CLONE

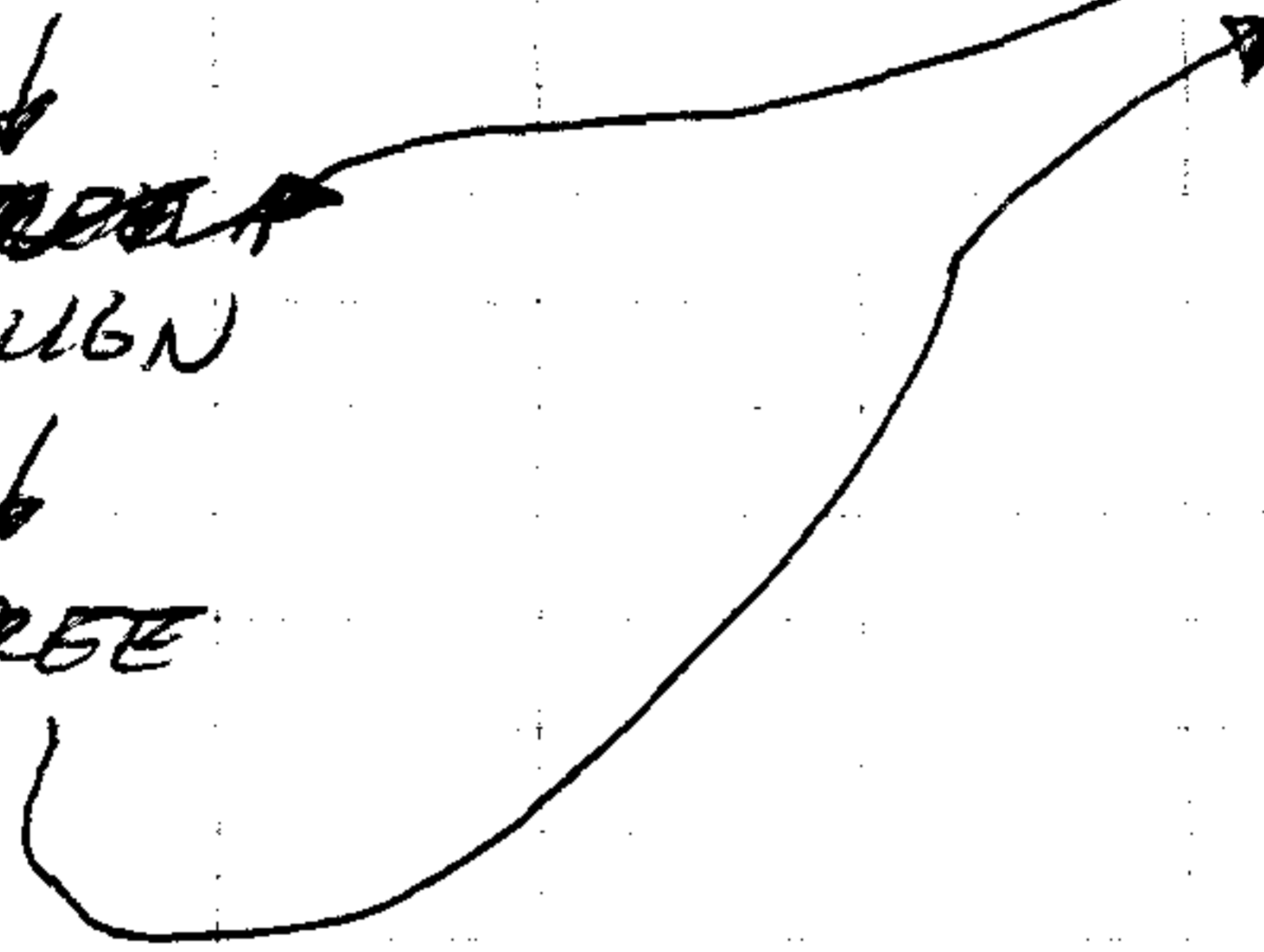
↓
SEQUENCE PART

→ SEQUENCE NOTE

↓
~~TRIAL~~

ALIGN

↓
TREE



I) Why interested in microbial diversity?

II) Why microbial diversity difficult?

A) Unculturable

B) Phenotype not meaningful

III) So - - molecular phylogenetics

A) Revolutionized view of microbial evolution

~~What is the structure of the tree?~~

① TREE -

② ARCHAEA -

③ ROW - & OTHER UNCULTURABLE

B) So - - how make tree

① RNA - translation

② conserved 2ary - ALIGN

③ signature sequences - PROBES

④ PCR easy

⑤ RATE GOOD

⑥ Free

⑦ RDP*

C) TREE

PARSIMONY, DISTANCE, ML

TREE
OF
LIFE

FIG 1, 2
FIG 3 - X
FIG 4

FIG 5 - X
FIG 6
FIG 7

Olsen - Archaea, Archaea, Everywhere

① easy to overlook microbes

② count molecules ...

- DeLong et al - find lots of Archaea - 30% of 4M biomass

- so much that it's like overlooking 300 elephants
in 1 km²

- in terms of biodiversity like overlooking all eukaryotes

③ what are Archaea

- molecularly determined separate group of proks

- but thought to only be in v. harsh environments

- acidic hot springs

- hydrothermal vents

- saturated brines

④ previous studies

Fuhrman et al 1992 } widely distributed in oceans
DeLong 1992 }

⑤ physiology?

- two lineages in ocean ... see figure

① one branches off methanogen / halophile group

② one branches with thermophiles

⑥ other

① lots of microdiversity -

② lots of macrodiversity -

DeLong et al

Method

- ① collect H₂O
- ② filter different sizes
- ③ purify DNA
- ④ purify rRNA
- ⑤ PCR
- ⑥ probe w/ Archaeal & Eukaryal probes
- ⑦ clone & sequence parts
- ⑧ phylogeny
- ⑨ sequence now from some

Results

- ① 18-30% of total picoplankton rRNA was Archaeal
- ② see Table 1 for other areas
- ③ 14 clones
- ④ 9/14 similar to hyperthermophiles Crenarchaeota
- ⑤ 5/14 similar to Euryarchaeota
- ⑥ max. likelihood supports assignment --- see Tree

Discussion

- ① high abundance in deep, cold H₂O suggests Archaea important part of biosphere
- ② only few areas looked at
- ③ implications for Archaeal evolution
- ④ what about physiology

Barns et al

① Intro

- can't cultivate many microbes
- seq. based phylogeny
- isolate genes from environment
- can use as probes to look at morphotype/abundance

② Method

① DNA

- collect sediment from Mud Volcano area
 - isolate DNA
 - PCR - using euk & arch specific primers
 - clone
- 3 x 9 m pool
 - some boiling
 - sediment black
 - high iron

② Trees

- align to RDP
- check chimera

① De Soete LSADT

② Phylip NJ

③ Paup parsimony } Bootstrapped

④ fastDNAml ML

③ Results

① All in Crenarchaeas clade

② trees mostly similar

③ but PJP 27 & 78 signatures of both suggest NOT either

④ bootstraps mostly high

⑤ transversion only same results so unlikely base composition problem

Barns et al

Discussion

- ① Great diversity - probably didn't cover all
- ② Representatives of major clades
- ③ Some w/ no clade
- ④ much erratic behavior

Problems, Etc

- ① phylogenetic placement questionable for some
- ② are the sequences artifacts?
- ③ sampling bias (PCR, clones, ...)

Future

- ① use to characterize morphology
- ② other probes
- ③ more areas
- ④ bacteria?
- ⑤ eukaryotes?