

Gordon Research Conference on Molecular Evolution

**Colony Harbortown Marina Resort
Ventura, California
January 28 - February 2, 1996**

**William R. Atchley, Chair
Walter M. Fitch, Vice-Chair**

This Conference is supported by the Alfred Sloan Foundation, the National Science Foundation, the Army Research Office and the Center for Quantitative Genetics at North Carolina State University.

Program for Gordon Conference in Molecular Evolution

| <u>Session Speaker</u> | <u>Topic</u> | <u>Discussion Leader</u> | <u>Time</u> |
|---------------------------------------|---|--------------------------|--------------------|
| Sunday evening, January 28 | | | |
| Opening Session | | Bill Atchley | 7:30-7:45 |
| <i>Early Evolution</i> | | Bill Schopf | 7:45-8:00 |
| Ford Doolittle | Rooting the tree of life | | 8:00-8:45 |
| Masami Hasegawa | Origin and early evolution of eukaryotes | | 9:00-9:45 |
| Monday morning, January 29 | | | |
| <i>Genome and Organelle Evolution</i> | | Dick Hudson | 8:30-8:45 |
| Priscilla Tucker | Evolution of the Mammalian Y-chromosome | <i>photo</i> | 8:45-9:30 |
| Jeff Palmer | Transfer of organelle genes to the nucleus | | 9:45-10:30 |
| John Doebley | Group Photo | | 10:50-11:10 |
| | Molecular basis of morphological evolution | | 11:20-12:05 |
| Monday evening, January 29 | | | |
| <i>Statistical inference</i> | | Masatoshi Nei | 7:30-7:45 |
| Nick Goldman | Mathematical modelling of molecular evolution | | 7:45-8:30 |
| Wen-Hsiung Li | Bootstrap techniques in phylogenetic analyses | | 8:45-9:30 |
| Tuesday morning, January 30 | | | |
| <i>Evolution of development</i> | | Andy Clark | 8:45-9:00 |
| Rudy Raff | Link between development and evolution | | 9:00-9:45 |
| Diethard Tautz | Evolution of gene networks | | 10:00-10:45 |
| William Atchley | Evolution of helix-loop-helix transcription factors | | 11:20-12:05 |
| Tuesday evening, January 30 | | | |
| <i>Experimental phylogenetics</i> | | Walter Fitch | 7:30-7:45 |
| J J Bull | Experimental phylogenetics | | 7:45-8:45 |
| <i>Poster session</i> | | | 8:45-10:00 |
| Wednesday morning, January 31 | | | |
| <i>Molecular Population Genetics</i> | | Morris Goodman | 8:45-9:00 |
| Maryellen Ruvolo | Inferring primate evolution with DNA sequence data | | 9:00-9:45 |
| Charles Aquadro | Determinants of genomic diversity: impact of recombination | | 10:00-10:45 |
| Dennis Powers | Evolution of gene structure and expression in natural populations | | 11:20-12:05 |
| Wednesday evening, January 31 | | | |
| <i>Non-tree-like evolution</i> | | Roger Milkman | 7:30-7:45 |
| Walter Fitch | Reticulation in HIV evolution | | 7:45-8:30 |
| Andreas Dress | Phylogenetic networks: Theory, software and visualization | | 8:45-9:30 |
| Thursday morning, February 1 | | | |
| <i>Viral evolution</i> | | Margaret Kidwell | 8:45-9:00 |
| Paul Sharp | Origins and evolution of AIDS viruses | | 9:00-9:45 |
| Susan Wessler | Transposable elements and the evolution of gene expression | | 10:00-10:45 |
| Walter Gilbert | New Arguments for Old Introns | | 11:20-12:05 |
| Thursday evening, February 1 | | | |
| <i>Pattern and Function</i> | | Jeff Thorne | 7:30-7:45 |
| Nancy Maizels | Phylogeny by function: origin of tRNA is in replication | | 7:45-8:30 |
| Marcella McClure | Identifying patterns in distantly related proteins | | 8:45-9:30 |

There will be a 15 minute question and discussion period following each talk, In the mornings, there will be a coffee break from 11:00 - 11:20 am (between the second and third talks).

Molecular Evolution Gordon Conference

Bill Schopf

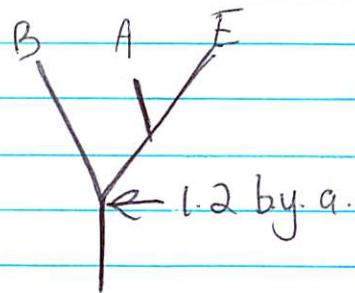
- R. Doolittle et al Science

suggest eukaryotic & bacterial common ancestor
was ~1.2 billion years ago

- why he doesn't think so

- Stromatolites ... were present

- cyanobacterial fossils



WF Doolittle - Constructive Evolution

- Complex & necessary interaction betw. molecules can be established by strictly neutral means

b - Evolutions big steps

- incr. in complexity → usu. given some adaptationist explanation.

Neutral alternative

- but no intrinsic reason for direction

- Complexity ... Maynard Smith & Szathmari 1995

- genome content α)

- cell types?

- body size

- interactions between molecules

possible measures
of complexity

2

THIS PROCESS IS DEPENDENT ON RELAXED SELECTION
FOR SIMPLICITY

- w RNA for example

why not more complex
in eubacteria

~~example~~

}

Neutral models for incr. in complexity

④ RNA editing

- C+U editing

- imagine there is a C+U deaminase that f(x) is on tRNA
- imagine it recognizes a site w/ a U
- then if there is a U>C mutation at this site
then these would be OK.

② Codon-capture & novel genetic codes Osawa - Evolution
of the genetic code

③ A. Lambowitz

- one intron reqs. Cyt-18 protein for splicing
- then other introns may be stabilized by prot-prot interaction or prot-tRNA
- then these may lose splicing ability

④ peptidyl-transferase

⑤ gene duplication



each become dependent on
diff. molecules

↓
↓

⑥ molecular drive

⑦ plasmid DNA • E. Zuckerkandl

M. Hasegawa - Origin & Evolution of Eukaryotes - Rooting the tree of life

Uncertainties in assumptions

MOLPHY 2.2

133,58 12.20 /pub/molphy*

Prot Ml

Advantages

- stochastic subst.
- robust (somewhat) to rate variation
- improves subst. model
- can evaluate total evidence from multiple genes

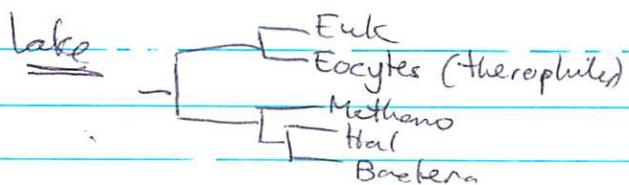
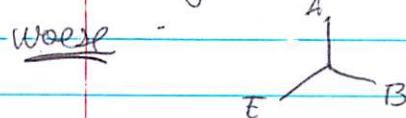
Amino-Acid substitution models

both assume
equilib. freq.
are average
freq.

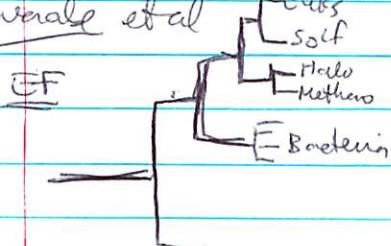
{ Dayhoff
Jones, Taylor, Thornton (CJTT)

can be improved by using actual eq freq. of each protein

Rooting tree of life



Inversible et al

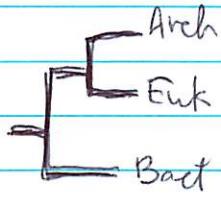


ATPase

showed sim. comparses
but due to paralogous comparsion

Brown & Dochtie

- AA tRNA synthetase
- Ile
- Val
- Leu

Early evolution of eukaryotes

phylogenetic placement of amitochondrial eukaryotes would be v. useful

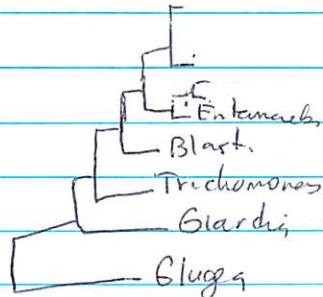
Problems w/ rRNA tree?

- GC content variation (e.g. GIARO1A ≈ 75%)

- ○ use protein sequences

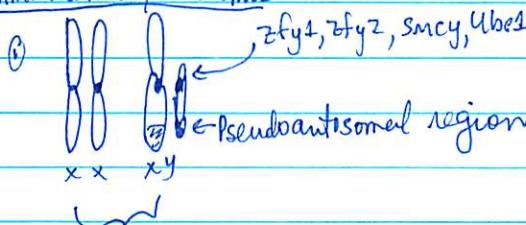
EF1- α

* - aa composition is not v. diff. from other EF1 α despite GC bias



✓ Priscilla Tucker - Evolution of sry (male sex determining locus)

Mammalian Sex Chromos



- only small region of homology

- so v. little recombination of Y chrom. specific genes

② appears to have evolved once in mammals

③ Y chromosome is late replicating

④ chromosome specific effects

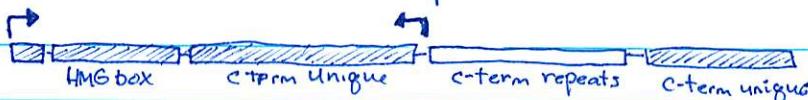
Other chrom. spec effects
① mutation

- ② hitch-hiking
 - ③ background selection
 - ④ Hill-Robertson effect
 - ⑤ bottleneck effects
 - ⑥ Muller's ratchet
 - ⑦ male-driven molecular evolution
- } more active in non-recombining regions

Sry

- involved in male tester development

SRY



- studied SRY in Murine genera

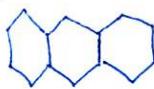
- what is variation w/in & betw. species?

- v. low variation in unique sequence regions

- multiple copies on Y in some species

- some variation among paralogs in C-term repeat regions

6



Explanation

GAG A G A

GAG A

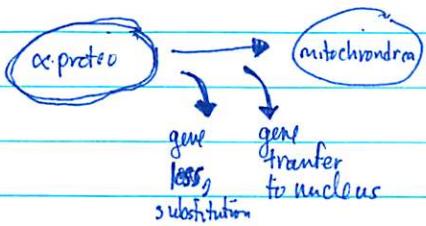


WY

Constructive & Destructive Neutral Evolution

J. Palmer - Transfer of Organellar Genes

Mitochondria



When did transfer occur?

- great variation in distribution / # of genes in diff. organelles

Gene Substitution

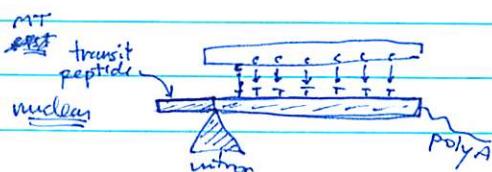
| | | | |
|-------------------------|----------|--------------|--------------|
| chloroplast transfer | p-tuta | glu synth | TAEV |
| | Fe-SO | Cu-Zn SO | |
| | F-1,6-BP | mal. dehydro | CPST |
| | GAPDH | TPI | substitution |
| | PGI | aldolase | |

Gene Transfer

- probably not still going on in yeast & animal mt DNA because of change in genetic code.

Recent Transfers?

- probe southerns w/ mt or CPST probes
- Cox II lost in some legumes nuclei
- many C → T changes in nuclear gene (the mt gene would be edited)
- most likely edited message was transferred
- some have it in nucleus, some in mt



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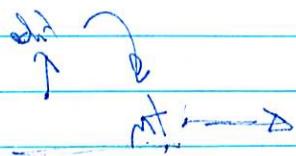
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How much is
THIS DUE TO
 $C \rightarrow T$ changes.

CoxII & Cob

- longer branch lengths in genes after transfer to nucleus

partitioning
C
↓
U



John Doeble - Molecular Basis of Morph. Evolution in Maize

Teosinte & Maize

- same biological species
- allows genetic analysis

Color

- teosinte doesn't have colored kernels, but does make anthocyanin
- Anthocyanin pathway
enzymatic vs. regulator genes

Tester crosses

- teosinte has dominant $f(x)$ alleles for all enzymatic loci

- R1 --- teosinte has recessive alleles \Rightarrow crosses w/ mutant have no kernel anthocyanin
- C1 --- teosinte also recessive

C1

- regulated by VP1 (VP1 activates C1)
- hypothesis
 - started w/ two indep. pathways
 - then these fused by VP1 activating C1 w/ acquisition of duplication in C1 promoter
 - but duplication thought to cause this was present in teosinte

10

Gene exchange?

HKA test -- nothing significant

C1.

- lowest Θ value for any maize locus

Paramutation

maybe due to methylation - one allele heritably alters another allele in heterozygotes at high frequency

Morphological Differences

QTL mapping



- many phenotypes mapped to long arm of chromosomes
 - submapped to region
 - this region contained teosinte branched gene
- used mutator line cross

mu x *tsb*

↓

mut
tsb or (μ) = mutant



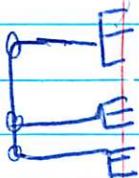
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100

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M.Nei

Reconstruction of Trees

- 1) Estimation of branch lengths $\&$ w/ topology
- 2) Estimation of topology



Rannala & Yang (manuscript)

~~estimator~~

- treats tree as random variable

- 3) Does a "sophisticated" substitution model improve phyl. reconstruction
- 4) Does the substitutional model remain the same over time

N. Goldman:

- what else is there in aligned sequences?

① Substitution matrices

② but

- diff. betw. codon positions
- diff. betw. positions



③ Codon-based models

New Stuff?

① Robustness (diff. models)

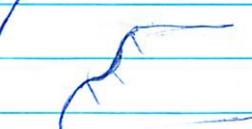
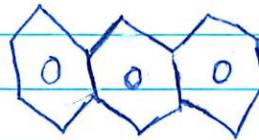
② Confidence

Testing Models

- ① Test a goodness of fit - are certain parameters important?
- transition - transversion?

Future things?

- ① invariant sites
- ② 2D structure
- ③ 3D structure
- ④ convergence & parallel changes
- ⑤ coding constraints



W-H Li - Bootstraps

Smaller sequences == more affected by variance w/
many parameters

Bootstrap

- what do % values mean

- boot % is less than prob. that tree is correct
- this underestimation incr. w/ more taxa

« with more taxa then less likely to get
a particular pattern by random »

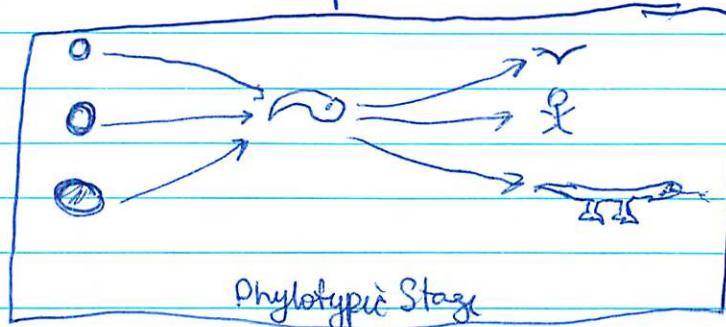
- the important parameter is # of alternative trees

A ClarkHomeotic genes

Evolutionary genetics of development
 Evolutionary of body plan

R. Raff

-Evolution of Development



- [] - suggests that the phylogenetic stage is the stage with the most protein-protein & other macromolecule interaction
 - o like WFD: "constructive evolution"

Sea Urchins

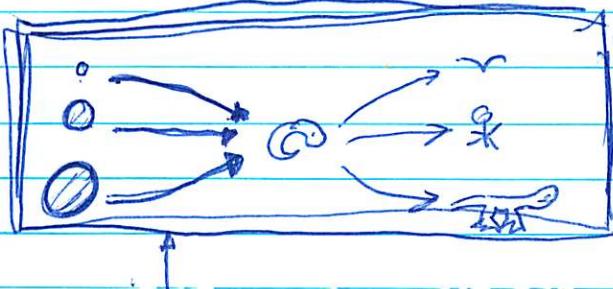
- two relatively closely related species
- start out devp. v. differently
- ~~evolution of phg~~

Evolution of phyla level body plan

D.Tautz - Evolution of Gene Networks (in embryos)

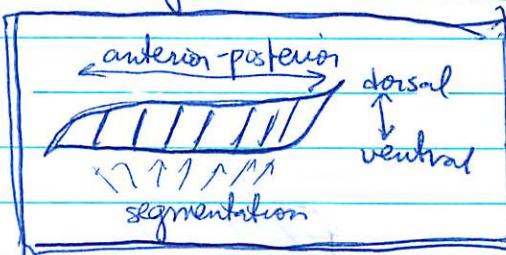
Molecular Comparative Embryology

- definition of homology may be diff. from other cases



- are the processes that lead to phylogenetic stage similar betw. species?

- bauplan genes
- segmentation genes → homeotic genes → cellular target genes



- embryo mostly tx factors → bauplan → cell fates

- the tx. factors form gradients - these work well in Drosophila embryo because all one cell essentially

- to remember

① lots of redundancy

② factors are expressed in phases

Many *hox* genes have similar $f(x)$ to gene homologs in other species

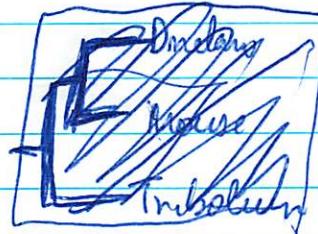
Regulatory gene networks appear to be highly conserved

But Not all genes retain their function

Depends on how you define $f(x)$.

FAST EVOLVING GENES

- ④ 100 randomly chosen embryonic cDNAs from *D. melanogaster*
 - ↓
 - hybridize to gDNA from flies (mosquito, housefly, beetles)
 - clone from close relative (*D. yakuba*)



Bill Atchley - Evolution of H2H proteins

Common tr. factor motifs

- Zn finger - leucine zippers
- HTH - steroid receptors
- H2H

H2H motif

- highly conserved: found in yeast → mammals
- often function in tr. activators
- examples

→ many many more

Myc, Max, Mad
MyoD
E12, ITF1
daughterless ...

- phylogenetic analysis

Cell Proliferation & Differentiation

Myc complex

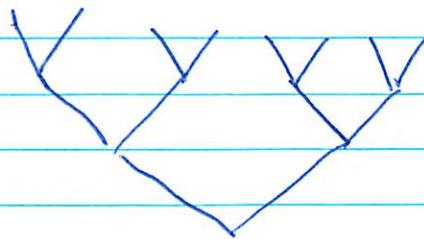
Jim Bull • Experimental Phylogenetics

Evolving bacteriophage in the lab

John H - never seen real data

i) Creating a Known phylogeny

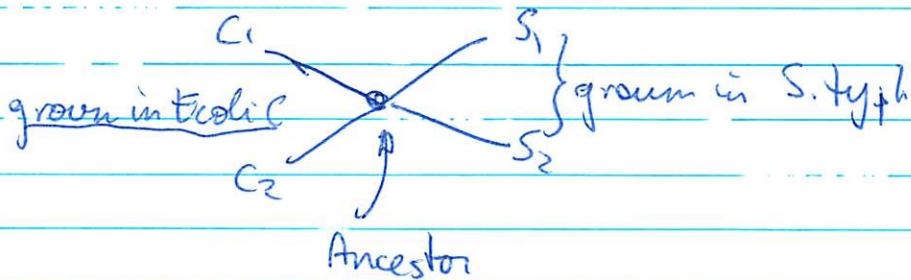
SHOULD
USE MIXED
POPULATIONS
AT NODES



- 1st phylogeny was symmetric
- grown in presence of mutagen
- restriction maps
- all methods gave right tree
- reconstruction of ancestral states v. good

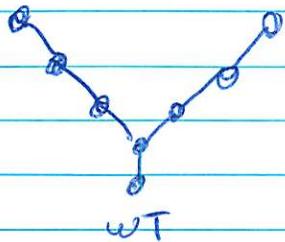
2) Convergent evolution in ϕ X174

- grown in phage chemo-stat
- adaptation to high T°



- 40% of sites that change were convergent

3) Convergence in T7



6x trees like this

- grown in mutagen
- in every phylogeny get specific deletion

4) Antisense

virus



add antisense



evolves resistance?



add new antisense

Morris Goodman - Globin Genes

Mary-Ellen Ruvo

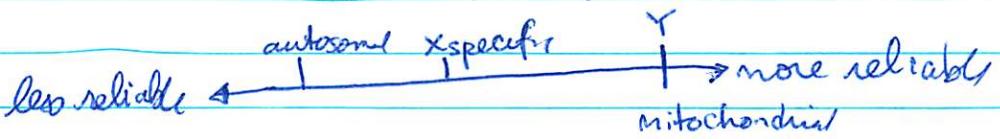
- When is an issue solved?

Using Multiple Genes

Single Gene Problems

① Ancestral polymorphism

- how likely are you to get gene-tree & species tree mismatches? see Takahata & Nei



Likelihood Ratio Method

- exclude some datasets

① those w/ trichotomy

② those w/ gene conversion

- can lead to lots of homoplasy

③ those w/ ancestral polymorphic

- combine closely linked genes

(Make sure to compare chimp, gorilla too)

Support Human-Chimp

- 11 gene sets

Support Gorilla-Chimp

- 2 data sets

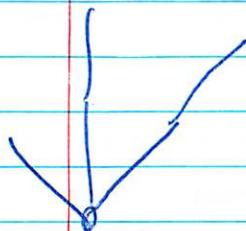
Support Gorilla-Human

- 1 data set

GCPDH

- Miyata et al 1987 (sperm vs. egg mutation)

If there
was a trichotomy.
plus uneven
branching



What about
recombination
being dependent
on heterozygosity

Do these models
assume recomb.
in a particular
region is constant
over time. Variation
might be important

Chip Aquadro

Recombination & Variation

At equilibrium:

- neutral = heterozygosity = $4N_e u$
- selection = reduces heterozyg.
- range of effect depends on recomb.

D. melanogaster

- can calculate rate of recomb. per physical unit
- highly variable
- levels of variability ^{w/in population} strongly correlated to recombination / physical unit

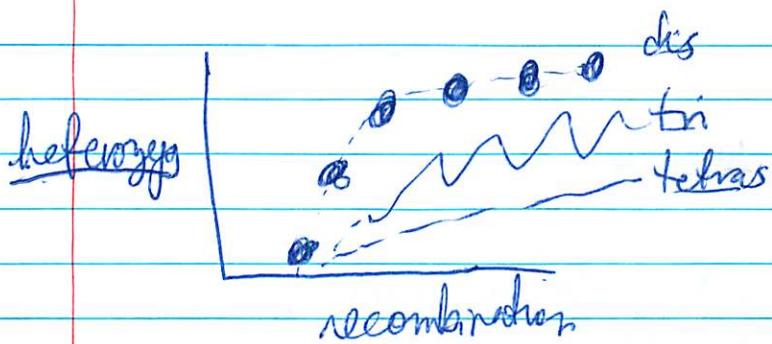
Explanations

- both predict [] you should have a correlation betw. recomb. & divergence betw. species
- ① genes in regions of low recomb. are f(x) constrained
 - ② recombination is mutagenic
 - ③ selective sweeps + hitchhiking
 - ④ background selection against deleterious mutations

[This is not seen]

Background Selection Model

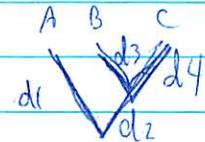
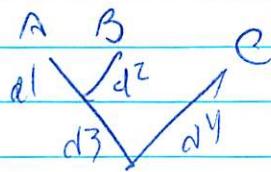
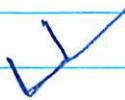
- only those regions free of deleterious mutations will ~~accumulate~~ persist in a population



Codon Bias } may also be affected by
GC recombination



L + x induces recomb



$$AC = d_1 + d_2 + d_4$$

$$AB = d_1 + d_2$$

$$BC = d_2 + d_3 + d_4$$

$$AC = d_1 + d_2 + d_4$$

$$AB = d_1 + d_2 + d_3$$

$$BC = d_3 + d_4$$

$A'C'$



WFitch - Networks

what to do if you have multiple equally "good" trees

① Consensus

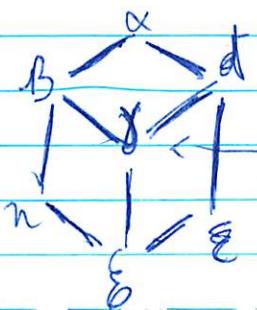
- strict-rule

- semi-strict → (keep those patterns that are not
contradictory w/ others)

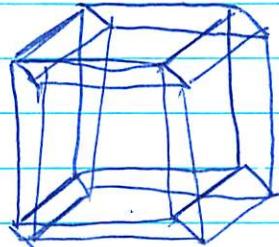
- majority rule

But consensus trees can be a problem:

- lose information



this sequence could arise
by multiple paths

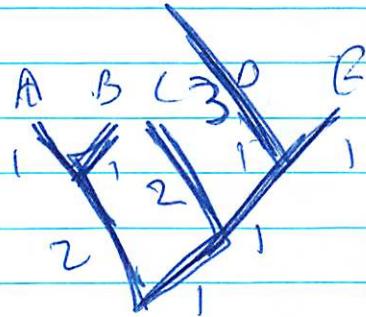


- sometimes this deals w/ parallel
changes and sometimes it
deals w/ ambiguity

- Networks can represent consensus trees
w/o including additional trees

4/19/11

| | | | | | | |
|---|---|---|---|---|---|-----|
| A | 0 | 2 | 6 | 6 | 6 | 5 |
| B | 2 | 0 | 6 | 6 | 6 | 5 |
| C | 6 | 6 | 0 | 4 | 4 | 5 |
| D | 6 | 6 | 4 | 0 | 2 | 4.5 |
| E | 6 | 6 | 4 | 2 | 0 | 4.5 |
| | A | B | C | D | B | |



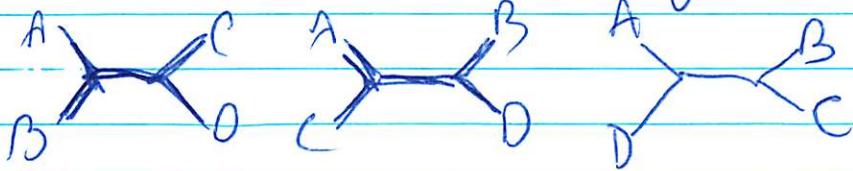
| | |
|---|---|
| A | 0 |
| B | 0 |
| C | 4 |
| D | 4 |
| E | 4 |

Networks - Split Decomposition

Represent non-tree-like patterns when they are then

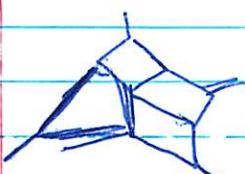
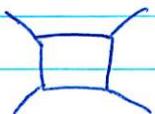
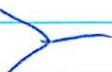
Buneman's Global Principle for trees

Suggest that w/ 4 taxa tree rather than trying to determine correct topology - to determine the least likely topology



Most methods would pick 1, 2, or 3.
He suggests rejecting 1, 2, or 3!

This is similar to Fourier analysis which decomposes periodic signals



sequence of speakers
 short interval
 long interval
 ↑ gilbert, late insertion
 M.Kidwell

Eukaryotic transposable elements

- ① Class I = retro-elements
- ② Class II = DNA elements



Link between retroelements & viruses

Evolution of TEs

Mode of evolution

- ① Ancient/modern?
- ② Phylogeny
- ③ Mode of transmission

Host-element relationships

- ① Strictly parasitic
- ② Mutualistic
- ③ Sequential parasitic/mutualistic

Gene conversion/recombination frequency?

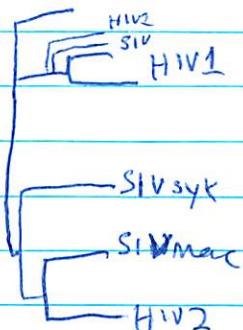
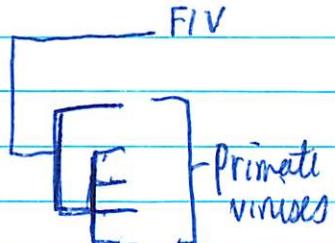
Host genes & cross-species transfer?

P. Sharp • Evolution of HIV

HIV evolution

- ① groups w/ Lentiviruses
 - ② w/ high rate
 - ③ appear to have been many cross-species transfers
 - ④ suggests no "good molecular clock" and that to get times of evolution of the virus to find regions of the tree w/ no cross-species events
- (is their a clustering of recombination in particular regions?)*

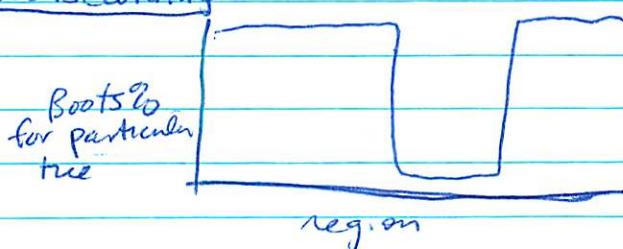
HIV2



Recombination in Retros/HIV

- compare phylogenies of diff. genes from same virus
- for this to occur an individual must be infected by two strains

Bootscanning



Sue Wessler

Are tRNA useful?

- McClintock suggested tRNAs were "controllers"

tRNA

Junkyard
Variety

Higher plants



- much of this is "junk"
- but how much is dead?

Two Genes

R = tx. activator

Waxy =

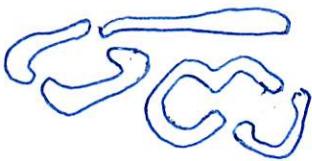
① Analyze mutant alleles

- how do tRNA's alter expression

② tRNA element families

Waxy Mutations

- isolate spontaneous mutation
- many insertions are stable because they do not encode tRNA and thus only stop when tRNA is expressed elsewhere
- many mutations / insertions also not have much phenotypic effect



Retrotransposons in Waxy

- many are spliced
- but not all the splicing is accurate
- some differences b/w different tissues

Ds elements transpose locally

TcNs affect genes by excision too

WaxK - jumped into waxy

- used as query sequence

- found many plant genes w/ remnants

only f Tourists (>10000 copies)

- similar to heartbreaker (1000s of copies - maybe?)

Slowaway - in monocots & dicots

Both prefer to insert in TA regions

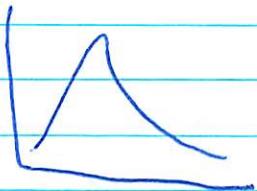
MITES

miniature-inverted-repeat-transposable-element

W. Gilbert

Exon shuffling - by more space ... get more recombination

Exon spectra



- appears that exons are reasonably correlated in size

Late
gain

Data

Phylogeny
- not in bacteria,
- but in vertebrates

Early

loss

introns at sim. positions

disagree w/ data

Correlation of introns
w/ 3D structure

agree w/ data

other stuff

- introns can be lost by rev. tx.

suggest correlation betw. modules & exons

- but if splicing is inaccurate then those insertion into ~~other~~ modules might be more deleterious

Intron Phase

- suggests that late insertion would not prefer any phase"

- suggests even shuffling favors in phase Ixony

but

① Selection agst. out of phase

② GC bias

③ Should get homologous introns in same position

Models

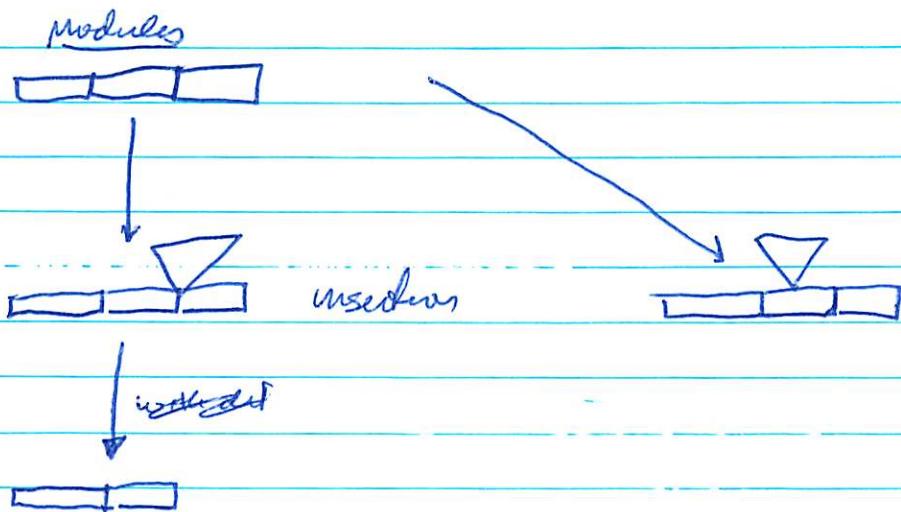
- are introns in linkers of proteins

nt

Size of Ixons 14 aa : 42

26 aa : 78

37 aa : 111



If splicing is not complete then the one at
~~at~~ ~~even~~ module border might be
less damaging.

Jeff ThorneAligning sequences

- evolutionary framework
- statistical basis
- feasible -

Using a tree to aid alignment

- Higgins
- Hein
- Feng & Doolittle
- Sankoff et al

Statistical basis

- Lloyd Allison

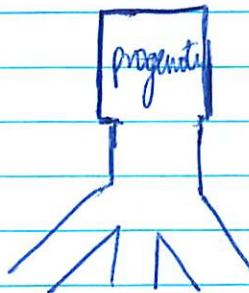
- integrate over all alignments allow each to contribute

- PNAS 91: 1059 JMB 235: 1501 Science 262: 208

Uses of evolution

- constraint identification
- multiple seq. align
- motifs
- structure predict
- database search

N. Margels



Molecular Fossils

- catalytic RNA

- suggests that those molecules that interact w/ lots of other molecules are MOST likely to remain as molecular fossils

(Now I'd like to go back to a simpler time)

RNA Genomes - Replication Problems

① Specificity?

② Telomere?

- circular

- special replication mechanism

- extra stuff at end that doesn't matter

- telomerase

- tag end

- e.g. Q_B-phage

5'

Q

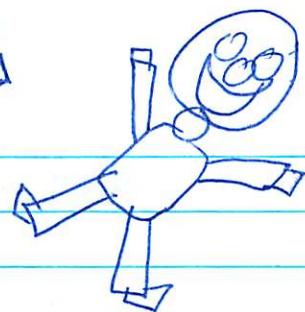
CCA-OH 3'

- replicase binds at end

- CCA works like telomere because there is a CCA adding enzyme

- also in mitochondrial plasmids

2/1/96
41



Contemporary tRNAs



added by CCA adding enzyme

RNA component
of RNase P cleaves
5' leader from
tRNA precursor



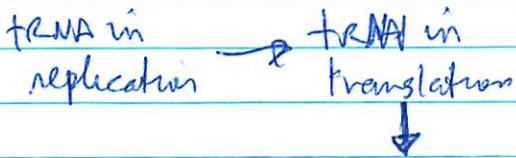
A. Leamboury

- Neurospora plasmid (Mauricenville)
- DS DNA plasmid
- replicate by rolling circle tx. w/ long RNA
w/ multiple copies

DNA
↓
multiple RNAs
↓
cleave
↓
rev. tx ass

How did tRNA get involved in translation

- suggests aas may have been used to change ^{term} structure to allow better priming.



- maybe aa tRNA synthetases evolved to change tRNA for replication
 ↓
 translation

May explain

① diversity of tRNA syn. structure

Suggests free tRNA may have evolved of two domain

CCA Adding enzyme

- some species add CCA

- some don't (CCA is already encoded)

M. McClure

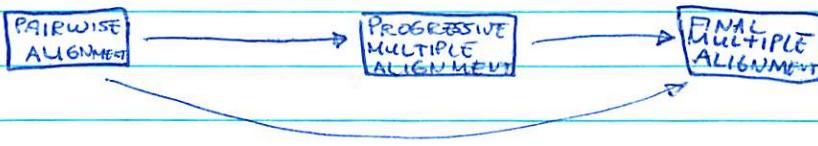
Reverse Transcriptase

- highly divergent

- needed better alignment algorithm

How well do multiple alignment programs do?

- every one uses pairwise alignment



Hidden Markov Model

- ordered series of motifs

- motif length

- Training Set Size

- Distribution of Similarity in training set

- Observed frequency of aa



dUTPase

- only found in two distinct retro-elements
- OS sequences

